Interrelationship of Hormone Concentration, Hormonal Imprinting and Receptor Down-regulation in Tetrahymena

György CSABA and László KÖHIDAI

Department of Biology, Semmelweis Uniwersity of Medicine, H-1445, Budapest, Hungary

Received on 5th May 1988, revised on 24th June 1988

Synopsis. Higher concentrations $(10^{-8}\text{-}10^{-6}\text{M})$ of insulin down regulated the insulin binding sites of Tetrahymena one day after treatment, however concentrations of $10^{-6}\text{-}10^{-18}\text{M}$ caused imprinting with higher binding capacity. After one week imprinting had been observed as a consequence of treatment by each dose. The experiments demonstrate, that (1) very low (100 femtomole) insulin concentration is enough for provoking imprinting and (2) down regulation is a response to hormone excess already at the level of Tetrahymena.

In higher organisms the primary interaction between the still immature, but genetically encoded receptor and the adequate hormone takes place in the perinatal period and gives rise to hormonal imprinting, which accounts for receptor amplification, i.e., for establishment of the hormone binding capacity characteristic of adulthood (C s a b a 1980, 1986 a). Since unicellular organisms are, by nature, lacking genetically encoded hormone receptors, they present membrane-associated protein configurations of varied quality as hormone binding structures, and transmit these to many progeny generations (C s a b a 1985, 1986 b, C s a b a et al. 1982). The establishment of such receptor "memory" is being influenced by many factors, of which the concentration of the hormone was the subject of the present study.

Materials and Methods'

Tetrahymena pyriformis GL cells maintained in 1% peptone plus 0.1 yeast extract containing Bacto-tryptone medium (Difco, Michigan, U.S.A.) at 28°C were treated (or not treated, as control) with different insulin (Semilente MC, Novo,

Denmark) concentrations (range: 10^{-8} - 10^{-18} M) for 1 h (sample size: 10 ml, volume 4×10^5 cell/ml), returned to plain medium for 24 h, then incubated in presence of fluoresceine-isothiocyanate-labeled insulin (FITC, BDH, England) for 1 h, and assayed for intensity of fluorescence in a Zeiss Fluoval cytofluorimeter, which was connected with a HP41CX calculator, programmed for the statistical evaluation of mean values, standard deviation, and significance of inter-group variation. At three levels of treatment (10^{-18} , 10^{-8} and 10^{-8} M) the fluorescence assay was done one week later, too. Twenty cells were assayed at each level of treatment in five replica experiments, thus the values shown in Fig. 1 represent means for 100 cells.

Results and Discussion

The experimental results have demonstrated that already a very low (100 fmol) concentration of insulin was sufficient to induce a lasting imprinting, which did not diminish after transmission to as many as 70 generations within a week's time. It was also demonstrated that insulin imprinting did not become stronger with the concentration increase, and exposure to higher insulin concentrations (10⁻³10⁻⁵M) even gave rise to down-regulation within 24 h (Fig. 1). This seems logical,

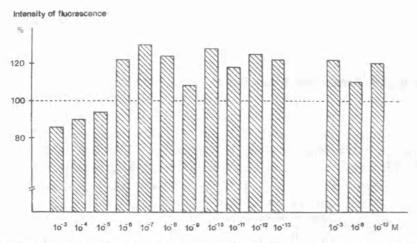


Fig. 1. Binding of insulin to Tetrahymena imprinted by different concentrations of insulin one day (left side) and one week (right side) after treatment, related to the control as 100%

if the appearance of coated pits and internalization — in every respect similar to those occurring in higher organisms — in Tetrahymena is taken into consideration (C s a b a et al. 1984). The critical threshold between the imprinting and down-regulatory effects of insulin on the unicellular is to be sought within the concentration range 10^{-5} - 10^{-6} M

but, after one week, the down-regulation elicited by high insulin level also transformed to an imprinting effect. Since, in view of this, imprinting was apparently independent of the applied hormone concentration, the presence, or rather the quality of the hormone seemed to play the decisive role. Since the "recognition" of, or "memory" of, useful and noxious environmental molecules is of vital importance for the survival and reproduction of a protozoon, the dependence of the imprinting mechanism on qualitative rather than quantitative factors seems to be wholly justified from the biological point of view.

Analysis of the present results from the angle of down-regulation a mechanism which has been well established in higher organisms (G a mmeltoft 1984, Gorden et al. 1980, Marshall and Olefsky 1980) — permits the conclusion that such a mechanism also operates at the unicellular level, at which it would also render the target cell refractory to excessive hormonal influence. Since down-regulation begins to operate already at the primary interaction of the target cell with the hormone if the latter is present in excess, there is reason to postulate that down-regulation is not so much an integral part of hormonal regulation as response shown to hormone excess also by "naive" - not encoded — cells after presentation of a hormone binding (receptor) structure (Csaba and Kőhidai 1986). The membrane of Tetrahymena being extraordinarily dynamic, exposure to insulin (or another hormone) for 1h seems to be sufficient to initiate either down-regulation or hormonal imprinting, depending on the applied concentration of the hormone.

REFERENCES

Csaba G. 1980: Phylogeny and ontogeny of hormone receptors: the selection theory of receptor formation and hormonal imprinting. Biol. Rev., 55, 47-63. Csaba G. 1985: The unicellular *Tetrahymena* as a model cell for receptor re-

search. Int. Rev. Cytol., 95, 327-377. Csaba G. 1986 a: Why do hormone receptors arise? Experientia, 42, 715-718. Csaba G. 1986 b: Receptor ontogeny and hormonal imprinting. Experientia, 42, 750-759.

Csaba G., Németh G. and Vargha P. 1982: Development and persistence of receptor "memory" in a unicellular model system. Exp. Cell. Biol., 50, 291-294.

Csaba G., Gruszczyńska M., Madarász B. and Sudár F. 1984: Coated pits and coated vesicles in unicellulars (Tetrahymena, Critidia) and in Hydra. Acta Morph. Hung., 32, 181-186. Csaba G. and Köhidai L. 1986: Modelling the insulin receptor in the *Tetra*-

hymena. Time dependence of receptor formation, down-regulation and imprinting. Acta Protozool., 25, 331-338.

Gammeltoft S. 1984: Insulin receptors: binding kinetics and structure-function relationship of insulin. Physiol. Rev., 64, 1321-1378.

Gorden P., Carpentier J.-L., Freychet P. and Orci L. 1980: Morphologic probes of polypeptide hormone receptor interactions. J. Histochem. Cytochem., 28, 811-817.

Marshall S. and Olefsky J. M. 1980: Effects of insulin incubation on insulin binding, glucose transport and insulin degradation by isolated rat adipocytes. J. Clin. Invest., 66, 763-772.